

Mini Review

DC-STAMP and Osteoimmunology

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The investigations from the aspect of “osteoimmunology” provided a recent advance on bone cell biology¹. Various molecules identified to play a crucial role in immune system also play a role in bone system. For example, RANK-RANKL system was originally identified in dendritic cells and T cells², but these molecules are both essential for osteoclast differentiation and skeletal development^{3,4}. Nuclear factor of activated T cells (NFATc1) was also originally identified in T cells, but plays an essential role in osteoclast differentiation⁵. Thus bone and immune systems share common molecules. Here we introduce a seven transmembrane receptor protein, dendritic cell specific transmembrane protein (DC-STAMP) as an “osteoimmunology” molecule, which plays a role in both bone and immune systems.

Rec.8/27/2008, 10/7/2008, pp59-62

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Key words DC-STAMP, osteoimmunology, osteoclasts, dendritic cells, cell-cell fusion

DC-STAMP and Bone system

Osteoclasts are bone-resorbing giant cells derived from hematopoietic stem cells or monocyte/macrophage lineage cells⁶. The most characteristic feature of osteoclasts is multi-nucleation induced by a cell-cell fusion of mono-nuclear osteoclasts. Various molecules such as macrophage fusion receptor, E-Cadherin and meltrin- α had been reported involved in osteoclast or macrophage cell-cell fusion⁷⁻⁹, however, defects in cell-cell fusion of osteoclasts and macrophages had not been shown by using knockout mice of these genes.

To isolate an osteoclast cell-cell fusion molecule, we undertook DNA subtraction screen between multi-nuclear osteoclasts and mono-nuclear macrophages, and we identified dendritic cell specific transmembrane protein (DC-STAMP) as an osteoclast

specific molecule¹⁰. DC-STAMP was originally isolated from dendritic cells or IL-4 stimulated macrophages^{11,12}, and was reportedly involved in osteoclast differentiation¹³. We generated DC-STAMP-deficient mice and found that DC-STAMP was essential for cell-cell fusion of osteoclasts and macrophages¹⁰. Cathepsin K, a terminal differentiation marker of osteoclasts, or c-Fos and NFATc1, both of which are essential transcription factors, were equally expressed in DC-STAMP-deficient and wild-type osteoclasts. The defect of cell-cell fusion in DC-STAMP-deficient osteoclasts was rescued by a forced expression of DC-STAMP. Thus DC-STAMP was considered specifically required for cell-cell fusion of osteoclasts rather than differentiation.

Although the multi-nucleation of osteoclasts is well known phenomena, the role of cell-cell fusion on bone system had not

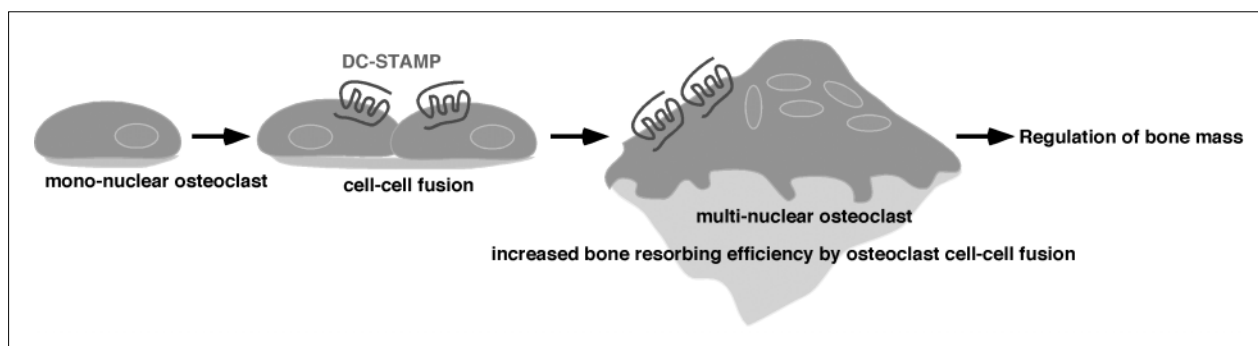


Fig.1 Osteoclast multi-nucleation and function

been characterized. We found that cell-cell fusion of osteoclasts upregulated the efficiency of bone resorption, and therefore, DC-STAMP-deficient mono-nuclear osteoclasts were less efficient to resorb bone compared with wild-type multi-nuclear osteoclasts. This leads DC-STAMP-deficient mice to increased bone mass than wild-type mice. Thus DC-STAMP regulates osteoclast cell-cell fusion, efficiency of osteoclast bone-resorption and physiological bone mass (Fig.1). Since DC-STAMP expression is directly regulated by c-Fos and NFATc1 in osteoclasts¹⁴, DC-STAMP plays a critical role in regulating bone homeostasis through osteoclasts. Recently, v-ATPase V0 subunit d2 (ATPv0d2) was reportedly involved in osteoclast cell-cell fusion¹⁵, and its expression was promoted by NFATc1¹⁶. The expression of ATPv0d2 was not different between DC-STAMP-deficient and wild-type osteoclasts¹⁵, suggesting that DC-STAMP and ATPv0d2 regulate osteoclast cell-cell fusion independently. Further studies are required to elucidate the mechanisms of osteoclast cell-cell fusion.

DC-STAMP and Immune system

Dendritic cells (DCs) are professional antigen presenting cells that present processed antigens to T cells. DC-STAMP was originally identified in dendritic cells¹¹, however, its specific role in DCs was not demonstrated. Since DCs do not fuse, it was speculated that DC-STAMP has a role other than cell-cell fusion in dendritic cells. We found that DC-STAMP regulated antigen presentation activity in DCs and maintenance of immune self-tolerance¹⁷.

DCs take up and process antigens, and present them to T cells in the context of MHC class I and II molecules. We found that antigen presentation activity of DC-STAMP-deficient DCs was higher than that of wild-type DCs in both class I and II pathways¹⁷. The lack of DC-STAMP did not inhibit differentiation or proliferation of DCs, suggesting that DC-STAMP specifically regu-

lates antigen presentation activity of DCs. Since the activation of T cells by DCs in both class I and II pathways is high in DC-STAMP-deficient DCs than wild-type DCs, a general mechanism, such as phagocytotic activity, may underlie high antigen presentation to class I and II pathways in DCs from DC-STAMP deficient mice. Indeed, DC-STAMP-deficient DCs showed higher phagocytotic activity than wild-type DCs.

Interestingly, aged DC-STAMP-deficient mice showed several systemic autoimmune symptoms including spontaneous lymphoproliferation, splenomegaly associated with infiltration of T-cells in several organs and increased serum anti-double stranded DNA antibody production¹⁷. Excessive immunization with activated DCs induces autoimmune disease^{18,19}, whereas, prolonged DC survival by inhibiting apoptosis of DCs induces autoimmunity²⁰, suggesting that over or prolonged antigen presentation by DCs induces autoimmune symptoms. DC-STAMP plays a role in regulating maintenance of immune self-tolerance through controlling antigen presentation of DCs.

Discussion

To date, various molecules have been identified to play a role in regulating both bone and immune systems. We previously demonstrated that osteoclasts and DCs share common precursor cells²¹. From common precursor cells, osteoclasts and DCs differentiate into professional bone resorbing and antigen presenting cells, respectively. DC-STAMP is highly expressed in both cells, and plays an essential role in regulating cell-cell fusion and bone resorbing efficiency in osteoclasts or phagocytosis and antigen presentation activity in DCs, respectively (Fig.2). Thus DC-STAMP plays a critical role in regulating both bone and immune system as an “osteoimmunology” molecule.

References

- 1) Takayanagi H: Osteoimmunology: shared mechanisms and

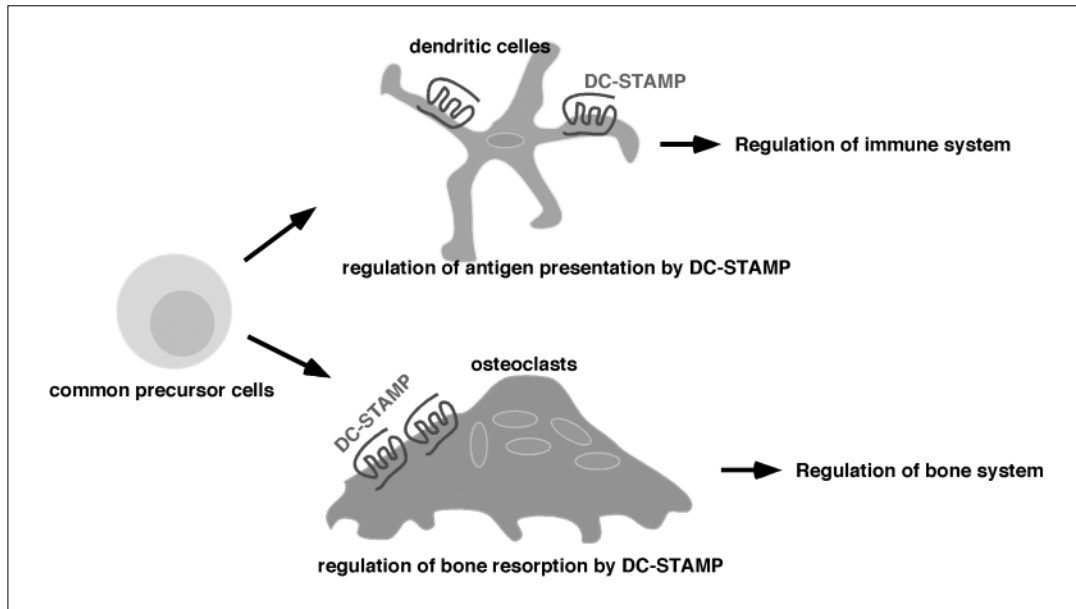


Fig.2 Role of DC-STAMP in bone and immune systems

- crosstalk between the immune and bone systems. *Nat Rev Immunol*, 7: 292-304, 2007.
- 2) Anderson DM, Maraskovsky E, Billingsley WL, Dougall WC, Tometsko ME, Roux ER, Teepe MC, DuBose RF, Cosman D, Galibert L: A homologue of the TNF receptor and its ligand enhance T-cell growth and dendritic-cell function. *Nature*, 390: 175-179, 1997.
 - 3) Kong YY, Yoshida H, Sarosi I, Tan HL, Timms E, Capparelli C, Morony S, Oliveira-dos-Santos AJ, Van G, Itie A, Khoo W, Wakeham A, Dunstan CR, Lacey DL, Mak TW, Boyle WJ, Penninger JM: OPG is a key regulator of osteoclastogenesis, lymphocyte development and lymph-node organogenesis. *Nature*, 397: 315-323, 1999.
 - 4) Dougall WC, Glaccum M, Charrier K, Rohrbach K, Brasel K, De Smedt T, Daro E, Smith J, Tometsko ME, Maliszewski CR, Armstrong A, Shen V, Bain S, Cosman D, Anderson D, Morrissey PJ, Peschon JJ, Schuh J: RANK is essential for osteoclast and lymph node development. *Genes Dev*, 13: 2412-2424, 1999.
 - 5) Takayanagi H, Kim S, Koga T, Nishina H, Isshiki M, Yoshida H, Saiura A, Isobe M, Yokochi T, Inoue J, Wagner EF, Mak TW, Kodama T, Taniguchi T: Induction and activation of the transcription factor NFATc1 (NFAT2) integrate RANKL signaling in terminal differentiation of osteoclasts. *Dev Cell*, 3: 889-901, 2002.
 - 6) Miyamoto T, Suda T: Differentiation and function of osteoclasts. *Keio J Med*, 52: 1-7, 2003.
 - 7) Saginario C, Sterling H, Beckers C, Kobayashi R, Solimena M, Ullu E, Vignery A: MFR, a putative receptor mediating the fusion of macrophages. *Mol Cell Biol*, 18: 6213-6223, 1998.
 - 8) Mbalaviele G, Chen H, Boyce BF, Mundy GR, Yoneda T: The role of cadherin in the generation of multinucleated osteoclasts from mononuclear precursors in murine marrow. *J Clin Invest*, 95: 2757-2765, 1995.
 - 9) Abe E, Mocharla H, Yamate T, Taguchi Y, Manolagas SC: Meltrin- α , a fusion protein involved in multinucleated giant cell and osteoclast formation. *Calcif Tissue Int*, 64: 508-515, 1999.
 - 10) Yagi M, Miyamoto T, Sawatani Y, Iwamoto K, Hosogane N, Fujita N, Morita K, Ninomiya K, Suzuki T, Miyamoto K, Oike Y, Takeya M, Toyama Y, Suda T: DC-STAMP is essential for cell-cell fusion in osteoclasts and foreign body giant cells. *J Exp Med*, 202: 345-351, 2005.
 - 11) Hartgers FC, Vissers JL, Looman MW, van Zoelen C, Huffine C, Figdor CG, Adema GJ: DC-STAMP, a novel multimembrane-spanning molecule preferentially expressed by dendritic cells. *Eur J Immunol*, 30: 3585-3590, 2000.
 - 12) Staeger H, Brauchlin A, Schoedon G, Schaffner A: Two novel genes FIND and LIND differentially expressed in deactivated and Listeria-infected human macrophages. *Immunogenetics*, 53: 105-113, 2001.
 - 13) Kukita T, Wada N, Kukita A, Kakimoto T, Sandra F, Toh K, Nagata K, Iijima T, Horiuchi M, Matsusaki H, Hieshima

- K, Yoshie O, Nomiyama H: RANKL-induced DC-STAMP is essential for osteoclastogenesis. *J Exp Med*, 200: 941-946, 2004.
- 14) Yagi M, Ninomiya K, Fujita N, Suzuki T, Iwasaki R, Morita K, Hosogane N, Matsuo K, Toyama Y, Suda T, Miyamoto T: Induction of DC-STAMP by alternative activation and downstream signaling mechanisms. *J Bone Miner Res*, 22: 992-1001, 2007.
- 15) Lee SH, Rho J, Jeong D, Sul JY, Kim T, Kim N, Kang JS, Miyamoto T, Suda T, Lee SK, Pignolo RJ, Koczon-Jaremkow B, Lorenzo J, Choi Y: v-ATPase V0 subunit d2-deficient mice exhibit impaired osteoclast fusion and increased bone formation. *Nat Med*, 12: 1403-1409, 2006.
- 16) Kim K, Lee SH, Ha Kim J, Choi Y, Kim N: NFATc1 induces osteoclast fusion via up-regulation of Atp6v0d2 and the dendritic cell-specific transmembrane protein (DC-STAMP). *Mol Endocrinol*, 22: 176-185, 2008.
- 17) Sawatani Y, Miyamoto T, Nagai S, Maruya M, Imai J, Miyamoto K, Fujita N, Ninomiya K, Suzuki T, Iwasaki R, Toyama Y, Shinohara M, Koyasu S, Suda T: The role of DC-STAMP in maintenance of immune tolerance through regulation of dendritic cell function. *Int Immunol*, 20: 1259-1268, 2008.
- 18) Ludewig B, Odermatt B, Landmann S, Hengartner H, Zinkernagel RM: Dendritic cells induce autoimmune diabetes and maintain disease via de novo formation of local lymphoid tissue. *J Exp Med*, 188: 1493-1501, 1998.
- 19) Roskrow MA, Dilloo D, Suzuki N, Zhong W, Rooney CM, Brenner MK: Autoimmune disease induced by dendritic cell immunization against leukemia. *Leuk Res.*, 23: 549-557, 1999.
- 20) Chen M, Wang YH, Wang Y, Huang L, Sandoval H, Liu YJ, Wang J: Dendritic cell apoptosis in the maintenance of immune tolerance. *Science*, 311: 1160-1164, 2006.
- 21) Miyamoto T, Ohneda O, Arai F, Iwamoto K, Okada S, Takagi K, Anderson DM, Suda T: Bifurcation of osteoclasts and dendritic cells from common progenitors. *Blood*, 98: 2544-2554, 2001.