

Mini Review

Influence of basic fibroblast growth factor in the solution and adsorbed form on the proliferation and differentiation of cells

Sachiko Inoue*, Yasuhiko Tabata

Department of Biomaterials, Field of Tissue Engineering, Institute for Frontier Medical Sciences, Kyoto University, Kyoto, Japan

There are some action modes to activate the receptor of growth factors and cytokines, such as juxtacrine, autocrine, and paracrine fashions. Some of growth factors transduce the signal to the target cell while anchored (immobilized or adsorbed) to a biological substance. For example, binding of fibroblast growth factor (FGF) to heparin is required for the signal transduction. Biological activity of growth factors, such as epidermal growth factor (EGF), nerve growth factor (NGF), brain-derived neurotrophic factor (BDNF), and insulin, in the immobilized or adsorbed form have been compared with that of factors in the water-soluble form. The growth factors immobilized or adsorbed maintain the activities to transduce the signals for activation of cell functions. The amount of growth factors immobilized to affect the proliferation of cells is smaller than that of water-soluble factors. There is difference in the time profile of the functional activation between the two forms of growth factor. This paper briefly reviews the influence of growth factors on the proliferation of cells while our data about the effect of basic FGF (bFGF) in different modes on the proliferation and differentiation of bone marrow derived stem cells.

Rec.1/10/2006, Acc.4/27/2006, pp181-184

* Correspondence should be address to:

Sachiko Inoue, Department of Biomaterials, Field of Tissue Engineering, Institute for Frontier Medical Sciences, Kyoto University, 53 Kawara-cho Shogoin, Sakyo-ku, Kyoto 606-8507, Japan. e-mail: yasuhiko@frontier.kyoto-u.ac.jp

Key words growth factor, immobilization, adsorption, solution, basic fibroblast growth factor

Introduction

Biological significance of growth factors has been increasingly magnified in terms of the medical researches and clinical applications. Many researches have clarified the receptor and signal transduction pathway of growth factors. Among of them, fibroblast growth factors (FGF) family consists of 18 types of FGF and binds 7 types of FGF receptors (FGFR). FGF-induced receptor dimerization is required for the signal transduction¹⁾.

Heparin is required for high affinity binding of basic FGF (bFGF) to the FGFR1 of cells that do not have an ability to synthesize cell-surface heparin sulfate^{2,3)}. The formation of heparin-induced acidic FGF (aFGF) dimer is the key step of FGFR2 dimerization⁴⁾. Therefore, the distance between FGF molecules and their anchoring to heparin are important to bind the FGFR and consequently activate the signal pathway (Fig.1a). Epidermal growth factor (EGF) and heparin-binding EGF-like growth fac-

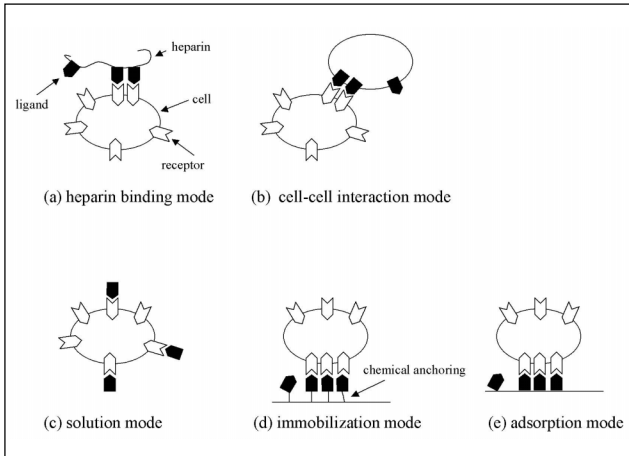


Fig.1 Modes of interaction between growth factor ligands and the receptors

tor (HB-EGF) bind to the epidermal growth factor receptor (EGFR) to activate cellular functions^{5,6}. It is reported that HB-EGF acts in the juxtacrine and paracrine fashions. HB-EGF of the former fashion is especially called proHB-EGF. ProHB-EGF is tethered to the plasma membrane of cells and can transmit the signal to the neighboring cell in contrast to the soluble mode (Fig.1b). The action of growth factors anchored to a biological substance or cell surface must be distinct from that of growth factors in the soluble form (Fig.1c).

Immobilization or adsorption and solution modes of growth factors

Growth factors are immobilized (Fig.1d) or adsorbed (Fig.1e) onto a solid substrate and the biological function has been compared with that of soluble growth factors (Fig.1c). Growth factor and cytokine immobilized or adsorbed on a solid substrate maintain their activity, and for some growth factor and cytokines, the time profile of their activation is different from that of water-soluble form. Chen et al. show that immobilization enabled EGF sufficiently to enhance the biological activity for cell growth of mouse STO fibroblasts and chinese hamster ovary cells (CHO)-K1⁷. Ito et al. have investigated the proliferation of rat pheochromocytoma cell line (PC12) cells and the activation of mitogen-activated protein kinase (MAPK) upon culturing in the presence of EGF in the immobilized and solution forms. Immobilized EGF activated the extracellular signal-regulated kinase (ERK) and p38 MAPK of cells for longer time periods than soluble EGF⁸. The MAPK activation induced by the immobilized EGF was observed in A431 cells and the activation lasted for a longer time period than that of soluble EGF⁹. Naka et al. have reported about the neurite outgrowth of PC12 cells cultured

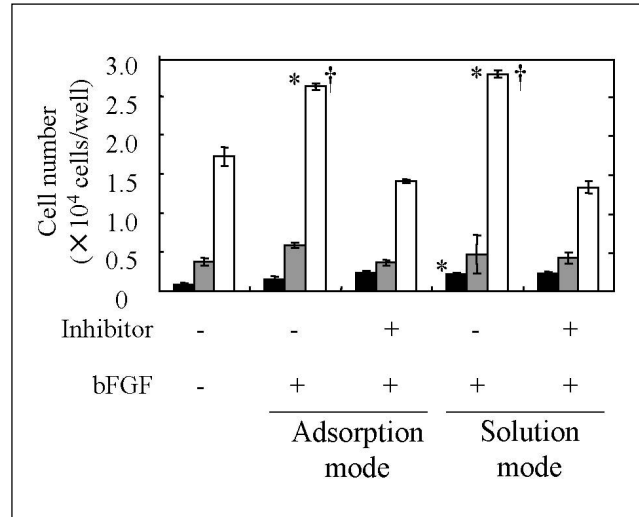


Fig.2 Proliferation profiles of BHK 21 cells cultured in the presence of bFGF adsorbed or soluble mode with or without the inhibitor for 3 (), 24(■), and 48() hrs. BHK 21 cells were incubated with 30 μ M of SU5402 for 5 min and seeded on cell culture plate (0.3 cm²) with bFGF in the adsorption (25 ng/cm²) and solution mode (100 ng/ml 75 μ l).

* $p < 0.05$: significant against the number of cells cultured without bFGF. † $p < 0.05$: significant against the number of cells cultures with bFGF after incubated inhibitor.

on magnetic beads with or without adsorption of nerve growth factor (NGF) or brain-derived neurotrophic factor (BDNF)¹⁰. The NGF or BDNF-adsorbed beads increased the neurite outgrowth of cells to the same extent as the original beads together with soluble NGF or BDNF. When the proliferation of CHO-T cells and activation of the insulin receptor, insulin receptor substrate 1, and phosphatidylinositol (PI)-3-kinase were investigated^{11,12}, both the insulin receptor phosphorylation and PI-3-kinase activity continued to increase over 12 hr for the insulin immobilized mode, in contrast to the soluble-mode insulin. The increased amount of total DNA was observed for insulin immobilized mode in contrast with the soluble insulin one.

There are some methods for artificial anchoring of growth factors. The anchoring methods were independent of the activity of growth factors and the growth factors induce the cell proliferation and differentiation. Irrespective of the immobilized mode, insulin at concentrations higher than 100 ng/cm² was effective in enhancing the cell proliferation¹¹⁻¹³. The immobilized and adsorbed of bone morphogenic protein-2 (BMP-2) increased alkaline phosphatase (ALP) activity of ST2 cells¹⁴.

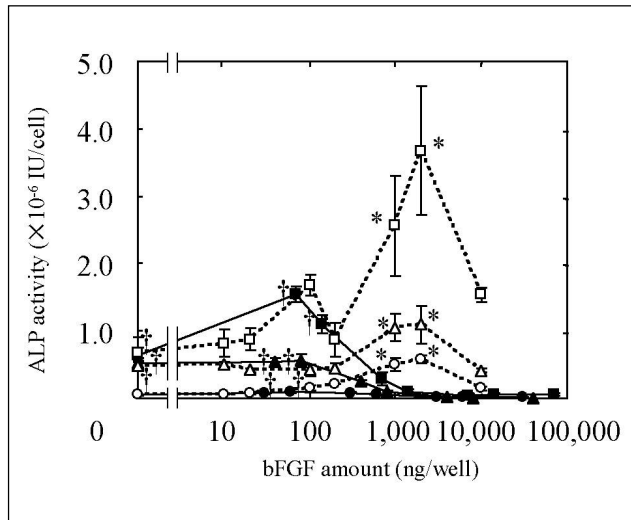


Fig.3 ALP activity of MSC cultured in osteogenic differentiation medium containing different amounts of bFGF for 10 (○, —), 14(□, ···), and 21(△, ···) days. bFGF was adsorbed onto the culture plates (2 cm², open symbols) or added to the medium (solid symbols).

* $p < 0.05$: significant against the ALP activity of MSC cultured in medium containing bFGF in adsorption form at 0, 10, 20 and 100 ng/well. † $p < 0.05$: significant against the ALP activity of MSC cultured in medium containing bFGF solution form at 1,000, 2,000, and 100,000 ng/well.

Adsorption and solution modes of bFGF

It has been well recognized that bFGF affects the proliferation and differentiation of various cells. There are some research reports on the addition effect of bFGF on the cell proliferation although little has been compared in the cell behavior between the addition modes. Baby hamster kidney (BHK) 21 cells have a FGFR and the proliferation is promoted by bFGF¹⁵. Fig.2 shows the proliferation profile of BHK21 cells cultured with bFGF in the adsorption or solution mode together with the addition effect of a FGFR1 inhibitor, SU5402¹⁶. Irrespective of the bFGF addition mode, bFGF presence increased the number of BHK 21 cells proliferated to a significant great extent compared with bFGF-free culture. A similar inhibition effect of SU5402 on the cell proliferation was observed for both the modes of bFGF. Taken together, it was found that bFGF adsorbed maintained the activity similar to that of free bFGF. BHK 21 cells have 60,000 sites of FGFR per cell to bind bFGF¹⁵. If we assume that cell spread-area is 600-1000 μm^2 and FGFR assemble in the side immobilized bFGF, BHK 21 cells would have FGFR at the density of 6×10^9 - 1×10^{10} receptor/cm². Three-dimensional structure of

bFGF was measured by x-ray. The cell dimension of bFGF crystals was about 3 nm¹⁷. If bFGF is adsorbed in a close packed state, the density of bFGF would be 1×10^{13} ligand/cm². bFGF adsorbed under our condition was 2.5 ng/cm², therefore 8×10^{11} ligand/cm² (18kDa)¹⁸. Therefore, when bFGF was adsorbed in a monolayer fashion, it is the number of ligand enough to bind the receptors of BHK 21 cells.

Fig.3 shows the level of ALP activity of rat bone marrow stromal cells (MSC) cultured in osteogenic differentiation medium containing bFGF in the adsorption or solution mode. The adsorption mode enabled MSC to significantly increase the ALP activity level compared with the solution mode. The level tended to increase with the amount of bFGF adsorbed, and attained to a maximum at the 10,000 ng/well over the time period studied. On the other hand, the opposite concentration dependence was observed for bFGF in the solution mode. The ALP activity decreased with the bFGF amount up to 1,000 ng/well, and thereafter became almost zero. Some researches have investigated the effect of bFGF addition into an osteogenic differentiation medium on the osteogenic differentiation of MSC. Locklin et al. revealed that the presence of bFGF enhanced the proliferation of MSC, but suppressed the osteogenic differentiation¹⁹. On the other hand, Frank et al. showed that bFGF induced BMP-2 expression²⁰. These findings indicate that osteogenic differentiation of MSC was influenced by the concentration of bFGF and the addition mode.

References

- 1) Mohammadi M, Olsen SK, Ibrahimi OA: Structural basis for fibroblast growth factor receptor activation. *Cytokine Growth factor Rev*, 16: 107-137, 2005.
- 2) Yayon A, Klagsbrun M, Esko JD, Leder P, Ornitz DM: Cell surface, heparin-like molecules are required for binding of basic fibroblast growth factor to its high affinity receptor. *Cell*, 64: 841-848, 1991.
- 3) Ornitz DM: FGFs, heparin sulfate and FGFRs: complex interactions essential for development. *BioEssays*, 22: 108-112, 2000.
- 4) Spivak-Kroizman T, Lemmon MA, Dikic I, Ladbury JE, Pinchasi D, Huang J, Jaye M, Crumley G, Schiessinger J, Lax I: Heparin-induced Oligomerization of FGF Molecules is responsible for FGF receptor dimerization, activation, and cell proliferation. *Cell*, 79: 1015-1024, 1994.
- 5) Higashiyama S, Lau K, Besner GE, Abraham JA, Klagsbrun M: Structure of heparin-binding EGF-like growth factor. *J Biol Chem*, 267: 6205-6212, 1992.
- 6) Singh AB, Harris RC: Autocrine, paracrine and juxtacrine signaling by EGFR ligands. *Cellular Signalling*, 17: 1183-

- 1193, 2005.
- 7) Chen G, Ito Y, Imanishi Y: Photo-immobilization of epidermal growth factor enhances its mitogenic effect by artificial juxtacrine signaling. *Biochim Biophys Acta*, 1358: 200-208, 1997.
- 8) Ito Y, Chen G, Imanishi Y, Morooka T, Nishida E, Okabayashi Y, Kasuga M: Differential control of cellular gene expression by diffusible and non-diffusible EGF. *J Biochem*, 129: 733-737, 2001.
- 9) Ogiwara K, Nagaoka M, Cho CS, Akaike T: Construction of a novel extracellular matrix using a new genetically engineered epidermal growth factor fused to IgG-Fc. *Biotechnol Lett*, 27: 1633-1637, 2005.
- 10) Naka Y, Kitazawa A, Akaishi Y, Shimizu N: Neurite outgrowths of neurons using neurotrophin-coated nanoscale magnetic beads. *J Biosci Bioeng*, 98: 348-352, 2004.
- 11) Ito Y, Chen G, Imanishi Y: Photoimmobilization of insulin onto polystyrene dishes for protein-free cell culture. *Biotechnol Prog*, 12: 700-702, 1996.
- 12) Ito Y, Zheng J, Imanishi Y, Yonezawa K, Kasuga M: Protein-free cell culture on an artificial substrate with covalently immobilized insulin. *Proc Natl Acad Sci USA*, 93: 3598-3601, 1996.
- 13) Hatakeyama H, Kikuchi A, Yamato M, Okano T: Influence of insulin immobilization to thermoresponsive culture surfaces on cell proliferation and thermally induced cell detachment. *Biomaterials*, 26: 5167-5176, 2005.
- 14) Tsujigiwa H, Nagatsuka H, Gunduz M, Rodriguez A, Rivera RS, LeGeros RZ, Inoue M, Nagai N: Effects of immobilized recombinant human bone morphogenetic protein-2/succinylated type I atelocollagen on cellular activity of ST2 cells. *J Biomed Mater Res*, 75A: 210-215, 2005.
- 15) Moscatelli D: High and low affinity binding sites for basic fibroblast growth factor on cultured cells: absence of a role for low affinity binding in the stimulation of plasminogen activator production by bovine capillary endothelial cells. *J Cell Physiol*, 131: 123-130, 1987.
- 16) Mohammadi M, McMahon G, Sun L, Tang C, Hirth P, Yen BY, Hubbard SR: Structure of the tyrosine kinase domain of fibroblast growth factor receptor in complex with inhibitors. *Science*, 276: 955-960, 1997.
- 17) Eriksson AE, Cousens LS, Weaver LH, Matthews BW: Three-dimensional structure of human basic fibroblast growth factor. *Proc Natl Acad Sci USA*, 88: 3441-3445, 1991.
- 18) Nugent MA, Iozzo RV: Fibroblast growth factor-2. *Int J Biochem Cell Biol*, 32: 115-120, 2000.
- 19) Locklin RM, Williamson MC, Beresford JN, Triffitt JT, Owen ME: In vitro effects of growth factors and dexamethasone on rat marrow stromal cells. *Clin Orthop Relat Res*, 313: 27-35, 1995.
- 20) Frank O, Heim M, Jakob M, Barbero A, Schafer D, Bendik I, Dick Walter, Hebberer M, Martin I: Real-time quantitative RT-PCR analysis of human bone marrow stromal cells during osteogenic differentiation in vitro. *J Cell Biochem*, 85: 737-746, 2002.