

Mini Review

Photo-damage mechanisms and anti-apoptotic effect of lutein in the mouse retina

Seiji Miyake^{1, 3)}, Mariko Sasaki^{1, 2)}, Noriko Takahashi^{1, 2)}, Kazuo Tsubota²⁾ and Yoko Ozawa^{1, 2, *)}

¹⁾Laboratory of Retinal Cell Biology, ²⁾Department of Ophthalmology, Keio University School of Medicine, Tokyo, Japan

³⁾Wakasa Seikatsu Co., Ltd., Kyoto, Japan

Photoreceptor cells receive light and transduce it to electrical signals for visual perception. However, excessive exposure to visible light causes photoreceptor cells to undergo apoptosis, which is called photo-damage. This damage involves several biochemical events, including the accumulation of oxidative stress and the elevation of intracellular calcium and nitric oxide (NO). Photo-damage is thought to be related to the progression of retinitis pigmentosa and age-related macular degeneration. Therefore, understanding the molecular mechanisms of retinal photodamage using model animals may lead to new therapeutic approaches for preventing the progression of these ocular diseases. In this review, we summarize previous reports examining the mechanisms of light-induced retinal damage, and briefly describe the interventional effect of lutein against photo-damage in mice. Lutein is taken from food and systemically delivered to the retina, skin, and certain organs and tissues. It reduces the level of reactive oxygen species and acts as an anti-oxidant in the retina of light-exposed mice, ultimately preventing light-induced DNA double-strand breaks and apoptosis. Although further study is required, lutein may be proposed as a new therapeutic approach for preventing photo-damage in humans.

Rec.3/29/2012, Acc.5/17/2012, pp208-212

*Correspondence should be addressed to:

Yoko Ozawa, M.D., Ph.D., Department of Ophthalmology, Keio University School of Medicine, 35 Shinanomachi, Shinjuku-ku, Tokyo 160-8582, Japan. Phone: +81-3-3353-1211, Fax: +81-3-3359-8302, E-mail: ozawa@a5.keio.jp

Key words light, oxidative stress, apoptosis, DNA double-strand breaks, retina

Introduction

Light is an essential external factor for living things, required for sight in animals as well as photosynthesis and growth in plants. Light is also important for developing visual system; it maturates, receiving light stimuli after birth¹). However, light can also induce adverse effects on the eyes; the exposure to excessive and/or intense light induces irreversible visual dysfunction. Noell et al. first demonstrated this effect in light-exposed animals²). That report was followed by extensive studies *in vivo* and *in vitro* on the relationship between light and retinal degeneration^{3, 4}).

Light-induced photoreceptor apoptosis is reported to occur in several phases, and many of the contributing factors have been identified^{4, 5}, although the entire process of

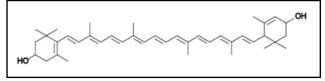


photo-damage has remained elusive. Interestingly, the first step (the induction phase) of the damage may be triggered by rhodopsin, an essential protein for light perception^{2, 6, 7}). For instance, rhodopsin knock-out mice are protected against photo-damage⁸⁾. Furthermore, inhibition of the visual cycle by 13-cis retinoic acid, a putative 11-cis retinal dehydrogenase inhibitor^{9, 10}, also prevents photo-damage¹¹. The chaperone protein RPE65 is distributed in the retinal pigment epithelium, where the photoreceptor cells undergo phagocytosis¹²); it is involved in the conversion of all-trans retinol, to 11-cis retinal¹³⁾. RPE65 knockout mice are also protected against light damage^{8, 14)}. Taken together, the evidence indicates that excessive stimulation of the visual cycle is an important mediator of photo-damage; moreover, accumulation of a rhodopsin bleaching intermediate, alltrans retinal, is now proposed to be responsible for photodamage in the retina¹⁵⁾.

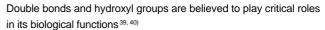
Following this induction phase, the death-signal phase can be divided into two sub-phases, early and late⁵). In the early phase, the intracellular calcium level increases, possibly caused by the activation of NO synthase. NO is a gaseous signaling molecule with physiological and pathological actions *in vivo*¹⁶). While a moderate level of NO in the central nervous system (CNS) is involved in synapse formation, its overexpression is reported to trigger intracellular disorders such as endoplasmic reticulum stress, mitochondrial morphologic change¹⁷, and membrane depolarization¹⁸.

In the late phase, AP-1 activation plays an essential role in mediating photoreceptor apoptosis¹⁷⁾. AP-1, a major nuclear transcription factor composed of c-Fos and c-Jun heterodimers, regulates various cellular events, including cell transformation, proliferation, differentiation, and apoptosis¹⁹⁾. Comprehensive gene expression analysis revealed that a component of the AP-1 transcription factor, c-Fos, is upregulated in the photo-damaged retina²⁰, and the DNAbinding activity of AP-1 is increased after light exposure²¹⁾. Mice that are deficient in c-fos exhibit normal retinal function and morphology²²⁾, but are highly resistant to photodamage, compared with wild-type mice²³⁾. Many current studies are aimed at understanding the role of AP-1 in retinal light damage; however, the molecules that function downstream of AP-1 activation in photo-damage are still unknown⁵⁾.

In addition to AP-1, caspases, a group of cysteine proteases²⁴, are also believed to contribute to retinal photo-







damage. In the light-exposed retina, the caspase-1 mRNA and protein levels increase^{25, 26)}, suggesting that at least caspase-1 participates in the induction of photoreceptor apoptosis.

An important implication of understanding the mechanism of photo-damage, is the possibility of developing new strategies for neuroprotection, in which these steps of the apoptotic pathway are inhibited. Several cytokines are reported to protect against photo-damage⁵⁾. Interestingly, a recent study revealed that erythropoietin, which stimulates hematopoiesis, exerts a neuroprotective effect on light-induced retinal degeneration²⁷⁾. An anti-inflammatory drug, naloxone, also reduces retinal damage^{28, 29)}, consistent with inflammatory events being associated with the light-exposed retina³⁰⁻³²⁾.

These observations led us to examine whether molecules with antioxidant activity could prevent retinal apoptosis and preserve visual function after light exposure. We recently showed that lutein (Fig.1), an antioxidant also known as a food factor, scavenges reactive oxygen species (ROS) and protects visual function against inflammatory ocular diseases^{33,34)}. Thus, we next evaluated the beneficial effect of the oral administration of lutein on light-induced retinal degeneration.

Lutein attenuates retinal photo-damage

To elucidate the protective effect of lutein on light-induced retinal degeneration, we exposed lutein-treated and vehicletreated BALB/c mice to 5000 lux of white light for 3 hours after at least 12 hours of dark adaptation. Five days after the light exposure, we performed electroretinography to assess the biological effect of lutein on visual function. In the vehicle-treated mice, light exposure induced a significant reduction in the amplitude of the a-wave, which reflects photoreceptor cell function, and the b-wave, which reflects the subsequent electrical reaction transmitted from the photoreceptor cells. However, strikingly, in the lutein-

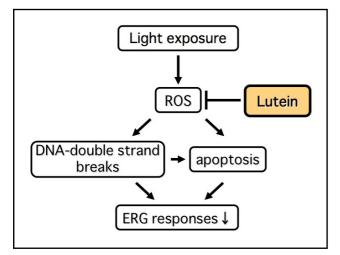


Fig.2 Protective effect of lutein against photo-damage in the retina

Lutein intake reduces the ROS level in the retina. This suppresses DNA-double strand breaks and apoptosis, finally preventing neuronal dysfunction.

treated mice, the reduction of these amplitudes was significantly attenuated. Because photoreceptor cells are susceptible to light exposure³⁵⁾, we also measured the thickness of the photoreceptor cell layer after light exposure. Consistent with the electroretinography results, the thickness of this cell layer was reduced after light exposure, and this effect was significantly suppressed in the luteinsupplemented mice compared with vehicle-treated mice.

One manifestation of apoptosis is the appearance of double-stranded DNA breaks (DSBs), which is detectable as DNA fragmentation⁴⁾. DNA damage results in the rapid phosphorylation of Histone H2AX, which has an important role in the repair of DSBs, at Ser139 in its C-terminus. To detect the effect of lutein in protecting against DSBs, the expression of phosphorylated H2AX (called gamma-H2AX) was examined by immunohistochemistry. The results showed that lutein-fed mice had fewer gamma-H2AX-positive photoreceptor cells than the controls.

The dephosphorylation of tyrosine142 of H2AX by EYA3 contributes to DNA repair rather than promoting apoptotic processes³⁶⁾. Therefore, we further investigated the expression of EYA3 with the concomitant detection of gamma-H2AX. EYA3 was expressed only in the photoreceptor cell layer after light exposure, and there were significantly more EYA3-expressing cells in the mice fed a lutein-supplemented diet than in control mice. This upregulation of EYA3-positive cells was also shown by western blotting.

To elucidate the effect of lutein on the ROS level in retinas after light exposure, the fluorescent probes dihydroethidium (DHE) and BODIPY-C11 were used as indicators of intracellular superoxide radicals³⁷⁾ and lipid peroxidation³⁸⁾, respectively. The fluorescence intensity of DHE increased in all the retinal layers after light exposure, but it was clearly suppressed in the mice fed the lutein-supplemented diet. The latter sign of oxidization appeared in the outer segment of photoreceptor cells in light-exposed mice fed control chow compared with non-light-exposed mice. However, this increase was significantly suppressed in the retinas of light-exposed mice fed a lutein-supplemented diet. These observations suggest that lutein's ROS-reducing effect may be protective against the terminal phase of photo-damage, reducing the amounts of DSBs and apoptosis (Fig.2).

Conclusion

The influence of light exposure on retinal damage increases with age, and is involved in the progression of some ocular diseases, such as retinitis pigmentosa (hereditary retinal degeneration) and age-related macular degeneration. Therefore, evidence-based preventive therapies against photo-damage are required. Photo-damage occurs not only in ocular tissues but also in the skin. Because lutein is physiologically obtained from food and delivered to the retina and skin, its therapeutic use against photodamage of both tissues may be feasible. Further studies aimed at revealing the molecular mechanisms of lutein's effects will help us discover new treatments for protecting tissues from photo-damage.

Aknowledgments

Some of our studies introduced in this mini-review were supported by a grant from Wakasa Seikatsu Co., Ltd., and in part by a grant-inaid from the Ministry of Education, Science, and Culture of Japan (MEXT) to Y.O. and M.S.

Seiji Miyake is employee of Wakasa Seikatsu Co., Ltd.

Conflict of interests

None

References

- 1) Wiesel TN, Raviola E: Myopia and eye enlargement after neonatal lid fusion in monkeys. Nature. 1977; 266: 66-68.
- 2) Noell WK, Walker VS, Kang BS, Berman S: Retinal

damage by light in rats. Invest Ophthalmol. 1966; 5: 450-473.

- 3) Hu D-N, Savage HE, Roberts JE: Uveal melanocytes, ocular pigment epithelium, and Müller cells in culture: in vitro toxicology. Int J Toxicol. 2002; 21: 465-472.
- Organisciak DT, Vaughan DK: Retinal light damage: mechanisms and protection. Prog Retin Eye Res. 2010; 29: 113-134.
- Wenzel A, Grimm C, Samardzija M, Remé CE: Molecular mechanisms of light-induced photoreceptor apoptosis and neuroprotection for retinal degeneration. Prog Retin Eye Res. 2005; 24: 275-306.
- Kaitz M, Auerbach E: Action spectrum for light-induced retinal degeneration in dystrophic rats. Vision Res. 1979; 19: 1041-1044.
- Williams TP, Howell WL: Action spectrum of retinal lightdamage in albino rats. Invest Ophthalmol Vis Sci. 1983; 24: 285-287.
- Humphries MM, Rancourt D, Farrar GJ, Kenna P, Hazel M, Bush RA, Sieving PA, Sheils DM, Creighton P, Erven A, Boros A, Gulya K, Capecchi MR, Humphries P: Retinopathy induced in mice by targeted disruption of the rhodopsin gene. Nature Genetics. 1997; 15: 216-219.
- 9) Law WC, Rando RR: The molecular basis of retinoic acid induced night blindness. Biochem Biophys Res Commun. 1989; 161: 825-829.
- 10) Gamble MV, Mata NL, Tsin A, Mertz JR, Blaner WS: Substrate specificities and 13-cis-retinoic acid inhibition of human, mouse and bovine cis-retinol dehydrogenases. Biochim Biophys Acta. 2000; 1476: 3-8.
- 11) Sieving PA, Chaudhry P, Kondo M, Provenzano M, Wu D, Carlson TJ, Bush RA, Thompson DA: Inhibition of the visual cycle in vivo by 13-cis retinoic acid protects from light damage and provides a mechanism for night blindness in isotretinoin therapy. Proc Natl Acad Sci USA. 2001; 98: 1835-1840.
- 12) Strauss O: The retinal pigment epithelium in visual function. Physiol Rev. 2005; 85: 845-881.
- 13) Saari JC: Biochemistry of visual pigment regeneration: the Friedenwald lecture. Invest Ophthalmol Vis Sci. 2000; 41: 337-348.
- 14)Grimm C, Wenzel a, Hafezi F, Yu S, Redmond TM, Remé CE: Protection of Rpe65-deficient mice identifies rhodopsin as a mediator of light-induced retinal degeneration. Nature Genetics. 2000; 25: 63-66.

- 15) Saari JC, Garwin GG, Van Hooser JP, Palczewski K: Reduction of all-*trans*-retinal limits regeneration of visual pigment in mice. Vision Res. 1998; 38: 1325-1333.
- 16) Ignarro LJ: Nitric Oxide, Second Edition: Biology and Pathobiology. Academic Press San Diego, USA; 2009. pp1-250.
- 17) Wenzel A, Grimm C, Marti A, Kueng-Hitz N, Hafezi F, Niemeyer G, Remé CE: c-fos controls the "private pathway" of light-induced apoptosis of retinal photoreceptors. J Neurosci. 2000; 20: 81-88.
- 18) Donovan M, Carmody RJ, Cotter TG: Light-induced photoreceptor apoptosis in vivo requires neuronal nitric-oxide synthase and guanylate cyclase activity and is caspase-3-independent. J Biol Chem. 2001; 276: 23000-23008.
- 19) Ameyar M, Wisniewska M, Weitzman JB: A role for AP-1 in apoptosis: the case for and against. Biochimie. 2003; 85: 747-752.
- 20) Chen L, Wu W, Dentchev T, Zeng Y, Wang J, Tsui I, Tobias JW, Bennett J, Baldwin D, Dunaief JL: Light damage induced changes in mouse retinal gene expression. Exp Eye Res. 2004; 79: 239-247.
- 21)Hafezi F, Marti a, Grimm C, Wenzel a, Remé CE: Differential DNA binding activities of the transcription factors AP-1 and Oct-1 during light-induced apoptosis of photoreceptors. Vision Res. 1999; 39: 2511-2518.
- 22) Kueng-Hitz N, Grimm C, Lansel N, Hafezi F, He L, Fox DA, Remé CE, Niemeyer G, Wenzel A: The retina of c-fos-/-mice: electrophysiologic, morphologic and biochemical aspects. Invest Ophthalmol Vis Sci. 2000; 41: 909-916.
- 23) Hafezi F, Steinbach JP, Marti A, Munz K, Wang ZQ, Wagner EF, Aguzzi A. Remé CE: The absence of cfos prevents light-induced apoptotic cell death of photoreceptors in retinal degeneration *in vivo*. Nature Medicine. 1997; 3: 346-349.
- 24)Kaufmann SH, Hengartner MO: Programmed cell death: alive and well in the new millennium. Trends Cell Biol. 2001; 11: 526-534.
- 25)Grimm C, Wenzel a, Hafezi F, Remé CE: Gene expression in the mouse retina: the effect of damaging light. Mol Vis. 2000; 6: 252-260.
- 26) Wu T, Chiang SKS, Chau FY, Tso MOM: Light-induced photoreceptor degeneration may involve the NF kappa B/caspase-1 pathway in vivo. Brain Res. 2003; 967: 19-26.

- 27) Grimm C, Wenzel A, Groszer M, Mayser H, Seeliger M, Samardzija M, Bauer C, Gassmann M, Remé CE: HIF-1-induced erythropoietin in the hypoxic retina protects against light-induced retinal degeneration. Nature Medicine. 2002; 8: 718-724.
- 28) Ni YQ, Xu GZ, Hu WZ, Shi L, Qin YW, Da CD: Neuroprotective effects of naloxone against light-induced photoreceptor degeneration through inhibiting retinal microglial activation. Invest Ophthalmol Vis Sci. 2008; 49: 2589-2598.
- 29) Yang L, Kim JH, Kovacs KD, Arroyo JG, Chen DF: Minocycline inhibition of photoreceptor degeneration. Arch Ophthalmol. 2009; 127: 1475-1480.
- 30)Ng TF, Streilein JW: Light-induced migration of retinal microglia into the subretinal space. Invest Ophthalmol Vis Sci. 2001; 42: 3301-3310.
- 31)Gordon WC, Casey DM, Lukiw WJ, Bazan NG: DNA damage and repair in light-induced photoreceptor degeneration. Invest Ophthalmol Vis Sci. 2002; 43: 3511-3521.
- 32) Rutar M, Provis JM, Valter K: Brief exposure to damaging light causes focal recruitment of macrophages, and long-term destabilization of photoreceptors in the albino rat retina. Curr Eye Res. 2010; 35: 631-643.
- 33) Sasaki M, Ozawa Y, Kurihara T, Noda K, Imamura Y, Kobayashi S, Ishida S, Tsubota K: Neuroprotective effect of an antioxidant, lutein, during retinal inflammation. Invest Ophthalmol Vis Sci. 2009; 50: 1433-1439.
- 34) Sasaki M, Ozawa Y, Kurihara T, Kubota S, Yuki K,

Noda K, Kobayashi S, Ishida S, Tsubota K: Neurodegenerative influence of oxidative stress in the retina of a murine model of diabetes. Diabetologia. 2010; 53: 971-979.

- 35) Kubota S, Kurihara T, Ebinuma M, Kubota M, Yuki K, Sasaki M, Noda K, Ozawa Y, Oike Y, Ishida S, Tsubota K: Resveratrol prevents light-induced retinal degeneration via suppressing activator protein-1 activation. Am J Pathol. 2010; 177: 1725-1731.
- 36) Cook PJ, Ju BG, Telese F, Wang X, Glass CK, Rosenfeld MG: Tyrosine dephosphorylation of H2AX modulates apoptosis and survival decisions. Nature. 2009; 458: 591-596.
- 37) Zhao H, Kalivendi S, Zhang H, Joseph J, Nithipatikom K, Vásquez-Vivar J, Kalyanaraman B: Superoxide reacts with hydroethidine but forms a fluorescent product that is distinctly different from ethidium: potential implications in intracellular fluorescence detection of superoxide. Free Radic Biol Med. 2003; 34: 1359-1368.
- 38) Drummen GP, van Liebergen LC, Op den Kamp JA, Post JA: C11-BODIPY581/591, an oxidation-sensitive fluorescent lipid peroxidation probe: (micro)spectroscopic characterization and validation of methodology. Free Radic Biol Med. 2002; 33: 473-490.
- 39) Ji HF: Insight into the strong antioxidant activity of deinoxanthin, a unique carotenoid in *deinococcus radiodurans*. Int J Mol Sci. 2010; 11: 4506-4510.
- 40) Johnson EJ: The role of carotenoids in human health. Nutr Clin Care. 2002; 5: 56-65.