Mini Review

Phospholipase C_{ε} as a potential molecular target for anti-inflammatory therapy and cancer prevention

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Phospholipase $C\varepsilon$ (PLC ε), encoded by PLCE1, is the phosphoinositide-specific PLC regulated by the ras proto-oncogene product Ras and its relative Rap1. Like other PLC isoforms, PLCε catalyzes hydrolysis of phosphatidylinositol 4,5-bisphosphate to yield second messengers, diacylglycerol and inositol 1,4,5trisphosphate. To understand the physiological function of PLC ε , we have created and analyzed genetically-modified mice in which PLC e is inactivated or overproduced. PLC e knockout (KO) mice are resistant to two-stage skin chemical carcinogenesis initiated by 7,12-dimethylbenz[a]anthracene and promoted by phorbol 12-myristate 13-acetate (PMA) and to intestinal tumorigenesis caused by the loss-of-function mutation of the APC tumor suppressor gene. In PLC KO mice, inflammation, such as that induced by a single application of PMA, that associated with tumorigenesis, and that in the elicitation phase of allergic contact hypersensitivity, is also attenuated as compared to that in wild-type mice. Conversely, overexpression of PLC ε in the epidermis results in the development of skin inflammation that partly shares the features with human psoriasis. In this article, we summarize the data showing the role of PLC in tumorigenesis and inflammation and discuss the future directions of the study of PLC ε for the development of therapies to control inflammation and to prevent cancer progression.

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Introduction

Phosphoinositide-specific phospholipase C (PLC) plays a pivotal role in regulation of intracellular signaling pathways. Upon activation of cell-surface receptors, such as serpentine receptors (also known as G protein-coupled receptors) and tyrosine kinase receptors, PLCs hydrolyzes phosphatidylinositol 4,5-bisphosphate (PIP2) into diacylglycerol (DAG) and inositol 1,4,5-trisphosphate (IP3). DAG binds to a variety of its target proteins including

Mutant mouse types	Experimental models	Phenotypes	References
Knockout	Two-stage skin chemical carcinogenesis (DMBA/PMA)	-Reduced tumor number -Inhibition of malignant progression	10)
	Intestinal tumorigeneis (<i>APC^{Min/+}</i> mice)	-Reduced tumor number -Inhibition of malignant progression -Decreased expression of proinflammatory molecules and angiogenic factors	15)
	Hapten-induced contact hypersensitivity (DNFB)	-Suppression of inflammation in elicitation phase -Reduced expression of proinflammatory molecule in the skin -No effect on the sensitization phase	12)
	PMA-induced skin inflammation	-Suppression of inflammation -Reduced expression of proinflammatory molecule in the skin	11)
Transgenic (skin-specific)		-Development of IL-23-dependent psoriasis-like dermatitis accompanied by aberrant infiltration of inflammatory cells, particularly those producing IL-2	

Table 1 Phenotypes of PLC ε mutant mice

protein kinase C (PKC) isoforms and Ras guanyl nucleotidereleasing proteins (RasGRPs) while IP3 stimulates the intracellular Ca²⁺ stores to release Ca²⁺ ion to the cytosol¹⁻⁴). In mammals, at least 13 PLC isoforms have been identified, and they are classified into six classes (β , γ , δ , ϵ , ζ , and η) based on the similarities in the structures and the mechanisms for the regulation¹⁻⁴).

PLC ε was first identified as a downstream target of the *ras* proto-oncogene product, Ras small GTPase⁵⁻⁶⁾. Subsequently, Rap1, another member of the Ras family of small GTPases, was also shown as an upstream molecule of PLC ε . PLC ε interacts with the GTP-bound active form of these small GTPases, and this interaction induces the translocation of PLC ε to the plasma membrane and the Golgi apparatus where PIP2 is hydrolyzed⁶⁻⁸⁾. In addition to Ras and Rap1, another small GTPase RhoA and heterotrimeric G proteins α_{12} and $\beta_{1\gamma_2}$ subunits are capable of activating PLC ε^{9} . In mammals, PLC ε is expressed in non-immune cells such as epidermal keratinocytes, dermal fibroblasts, and epithelial cells, but not in immune cells such as lymphocytes, granulocytes, macrophages, and dendritic cells¹⁰⁻¹³⁾.

In this mini-review article, we summarize the results obtained from the study of genetically-modified mice in which PLC ε loses its lipase activity or is overproduced (Table 1). We discuss the future directions of the study of PLC ε for the development of therapies that control inflammation and prevent cancer progression.

Tumorigenesis

To address the physiological role of PLC ε , PLC ε knockout mice were created by disrupting one of the exons encoding the domain responsible for its lipase catalytic activity¹⁴). The first evidence for the role of PLC ε in tumorigenesis was provided by the experiments employing the two-stage skin chemical carcinogenesis protocol using 7,12-dimethylbenz[a]anthracene (DMBA) and phorbol 12-myristate 13-acetate (PMA) as an initiator and a promoter, respectively¹⁰). PLC ε KO mice developed only a small number of benign tumors compared to wildtype (WT) mice, and their malignant progression into carcinomas appeared to be suppressed.

The second evidence was provided by the study using $APC^{Min/+}$

mice¹⁵⁾, which develop multiple intestinal neoplasms (Min) due to the loss-of-function mutation in the *APC* tumor suppressor gene¹⁶⁾. Intestinal tumors developed in PLC ε -deficient *APC*^{Min/+} mice were markedly decreased in number, and their malignant progression was strongly suppressed compared to those in PLC ε sufficient *APC*^{Min/+} mice. Notably, infiltration of inflammatory cells and expression of proinflammatory molecules (e.g., cyclooxygenase-2, chemokine (C-X-C motif) ligand (Cxcl)-1, and Cxcl-2) in the intestinal tumors of PLC ε -deficient *APC*^{Min/+} mice were considerably decreased compared to those in the gradematched tumors of PLC ε -sufficient *APC*^{Min/+} mice. These results suggest that the oncogenic effects of PLC ε are mediated by augmentation of inflammation.

Recent genome-wide studies have identified susceptibility loci of gastric cardia adenocarcinoma and esophageal squamous cell carcinoma in *PLCE1*, the gene encoding PLC $\varepsilon^{17,18}$. Although the impacts of the single nucleotide polymorphisms identified in these human cases are not understood, these findings suggest that PLC ε is involved in tumorigenesis also in humans.

PMA-induced skin inflammation and $PLC\varepsilon$ activation¹¹⁾

In $PLC\varepsilon^{\neq}$ mice, infiltration of leukocytes in the skin after a single application of PMA was substantially suppressed compared to that in $PLC\varepsilon^{\neq/+}$ mice. This alteration highly correlated with the attenuation of PMA-induced expression of interleukin (IL)-1 α in the skin. Such $PLC\varepsilon$ -dependent IL-1 α induction was reproduced in primary-cultured dermal fibroblasts. Similarly, Cxcl-2 induction after PMA treatment in cultured dermal fibroblasts was also found to be $PLC\varepsilon$ -dependent. These results suggested that PLC ε plays a role in PMA-triggered upregulation of cytokine expression in dermal fibroblasts.

The above-presented results also suggested that PLC ε is activated in dermal fibroblasts after PMA treatment. In order to examine this possibility, we assayed PLC activity in PMA-stimulated cells by measuring the cytosolic free Ca²⁺ concentration whose increase is mediated by the PLC product IP₃. In *PLC* ε ^{-/-} fibroblasts, the elevation of the cytosolic free Ca²⁺ concentration after PMA application was substantially attenuated as compared to that in *PLC* ε ^{+/+} fibroblasts, indicating that PMA activates PLC ε . As an upstream activator of PLC ε , Rap1 appears to play a major role because Rap1 was activated upon PMA treatment and the siRNA-mediated knockdown of Rap1 expression almost totally abolished PMA-induced PLC ε activation. The PMA-induced Rap1 activation requires RasGRP3, which is the DAG/ PMA-responsive Rap1 GEF, and PKCs (Fig.1).



Fig.1 PMA-induced activation of PLC ε PMA activates Rap1 through the DAG/PMA-responsive Rap1 GEF, RasGRP3, and PMA-responsive PKC. The GTP-bound active Rap1 in turn activates PLC ε . Oncogenically activated Ras may also be involved in PLC ε activation. Activation of PLC ε results in the production of proinflammatory molecules such as IL-1 α , causing inflammation and promoting tumor formation.

Allergic contact hypersensitivity (CHS)¹²⁾

In the elicitation phase of CHS induced by the hapten 2,4dinitrofluorobenzene (DNFB), PLC ε -deficiency attenuated the expression of proinflammatory cytokines by epidermal keratinocytes as well as dermal fibroblasts in vivo without interfering with the infiltration of CD4⁺ T cells in the hapten-painted sites. Adoptive transfer of CD4+ T cells from DNFB-sensitized $PLC\varepsilon^{-/-}$ mice could induce allergic CHS in naïve mice on the $PLC\varepsilon^{+/+}$ background. On the other hand, naïve mice receiving the hapten-primed CD4⁺ T cells on the PLC $\varepsilon^{-/-}$ background exhibited reduced CHS response as compared to those on the *PLC* $\varepsilon^{+/+}$ background (Fig.2). These results indicated that the elicitation phase is PLC ε -dependent while the sensitization phase does not require PLC ε at all. Experiments using primary-cultured cells demonstrated that both fibroblasts and keratinocytes express proinflammatory cytokine genes after stimulation with T-cell-derived cytokines such as IL-17, IL-4, tumor necrosis factor- α , and interferon- γ in a PLC ε -dependent manner. How-



Fig.2 Role of PLCε in the elicitation phase of CHS demonstrated by adoptive transfer of CD4⁺ T cells

CD4⁺ T cells were adoptively transferred form the DNFB-sensitized mice on the different *PLC* ε background to naïve mice on the different *PLC* ε background as illustrated. PLC ε -deficiency in the recipient mice attenuated CHS response.

ever, the T-cell-derived cytokines tested failed to function as a ligand that triggers the activation of PLC ε , suggesting that certain soluble factors secreted upon cytokine stimulation may mediate PLC ε activation leading to proinflammatory gene activation in a *PLC* ε -dependent manner.

Inflammation in PLC ϵ -overexpressing transgenic mice¹³⁾

Role of PLC ε in inflammation has been further proven with the transgenic mice overexpressing PLC ε in the epidermis using the tissue-specific expression system, in which Cre recombinase induces the expression of the transgene by removing the Cre-recognizable silencing sequences. Epidermis-restricted overexpression was achieved by expressing Cre under the control of the basal layer-specific keratin 5 promoter. The resulting mice spontaneously developed skin inflammation sharing features with human psoriasis, such as formation of adherent silvery scale, excessive growth of keratinocytes, and aberrant infiltration of immune cells to the skin. The development of the skin symptoms in the PLC ε transgenic mice was accompanied by increased production of IL-23 from keratinocytes and the accumulation of IL-22-producing CD4⁺ lymphocytes whose nature has not been fully characterized. IL-23 appeared to be crucial for the skin inflammation caused by PLC ε overexpression because intradermal injection of a neutralizing antibody against IL-23 totally reversed these symptoms. These results suggest a possible role of the PLC ε -mediated signaling in the pathogenesis of inflammatory diseases involving aberrant IL-23 production, such as psoriasis.

Discussion

As reviewed in this article, our studies using PLC & knockout mice have demonstrated that PLC ε plays an important role in regulation of proinflammatory molecule expression in non-immune cells such as keratinocytes and fibroblasts, thereby augmenting inflammation and probably promoting tumorigenesis. The data showing the skin inflammation in the PLC ε -overexpressing mice imply that deregulation of the PLCe-mediated signaling may be involved in the pathogenesis of chronic inflammatory diseases. These results suggest that PLC ε could be a molecular target for the therapies of inflammatory diseases and for the prevention of carcinogenesis. However, to develop the therapeutic methods targeting PLC ε , there are a number of problems to be solved. Which ligands trigger the activation of PLC ε in inflammatory processes? How does PLCe operate on inducing proinflammatory cytokine expression? Further studies are underway to address these questions.

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References

- Rhee SG: Regulation of phosphoinositide-specific phospholipase C. Annu Rev Biochem. 2001; 70: 281-312.
- Fukami K: Structure, regulation, and function of phospholipase C isozymes. J Biochem. 2002; 131: 293-299.
- Katan M: New insights into the families of PLC enzymes: looking back and going forward. Biochem J. 2005; 391: e7e9.
- 4) Suh PG, Park JI, Manzoli L, Cocco L, Peak JC, Katan M, Fukami K, Kataoka T, Yun S, Ryu SH: Multiple roles of phosphoinositide-specific phospholipase C isozymes. BMB

Rep. 2008; 41: 415-434.

- 5) Kelley GG, Reks SE, Ondrako JM, and Smrcka AV: Phospholipase C ε : a novel Ras effector. EMBO J. 2001; 20: 743-754.
- 6) Song C, Hu CD, Masago M, Kariyai K, Yamawaki-Kataoka Y, Shibatohge M, Wu D, Satoh T, Kataoka T: 2001. Regulation of a novel human phospholipase C, PLCε, through membrane targeting by Ras. J Biol Chem. 2001; 276: 2752-2757.
- Jin TG, Satoh T, Liao Y, Song C, Gao X, Kariya K, Hu CD, Kataoka T: Role of the CDC25 homology domain of phospholipase Cε in amplification of Rap1-dependent signaling. J Biol Chem. 2001; 276: 30301-30307.
- Song C, Satoh T, Edamatsu H, Wu D, Tadano M, Gao X, Kataoka T: Differential roles of Ras and Rap1 in growth factor-dependent activation of phospholipase C epsilon. Oncogene. 2002; 21: 8105-8113.
- Bunney TD, Katan M. Phospholipase Cε : linking second messengers and small GTPases. Trends Cell Biol. 2006; 16: 640-648.
- Bai Y, Edamatsu H, Maeda S, Saito H, Suzuki N, Satoh T, Kataoka T: Crucial role of phospholipase Cε in chemical carcinogen-induced skin tumor development. Cancer Res. 2004; 64: 8808-8810.
- Ikuta S, Edamatsu H, Li M, Hu L, Kataoka T: Crucial role of phospholipase Cε in skin inflammation induced by tumor-promoting phorbol ester. Cancer Res. 2008; 68: 64-72.
- 12) Hu L, Edamatsu H, Takenaka N, Ikuta S, Kataoka T: Crucial role of phospholipase Cε in induction of local skin inflammatory reactions in the elicitation stage of allergic contact hypersensitivity. J Immunol. 2010; 184: 993-1002.
- 13) Takenaka N, Edamatsu H, Suzuki N, Saito H, Inoue Y, Oka M, Hu L, Kataoka T: Overexpression of phospholipase Cε in keratinocytes upregulates cytokine expression and causes dermatitis with acanthosis and T-cell infiltration. Eur J Immunol. 2011; 41: 202-213.
- 14) Tadano M, Edamatsu H, Minamisawa S, Yokoyama U, Ishikawa Y, Suzuki N, Saito H, Wu D, Masago-Toda M, Yamawaki-Kataoka Y, Setsu T, Terashima T, Maeda S, Satoh T, Kataoka T: Congenital semilunar valvulogenesis defect in mice deficient in phospholipase Cε. Mol Cell Biol.

2005; 6: 2191-2199.

- 15) Li M, Edamatsu H, Kitazawa R, Kitazawa S, Kataoka T: Phospholipase C ε promotes intestinal tumorigenesis of $Apc^{Min/+}$ mice through augmentation of inflammation and angiogenesis. Carcinogenesis. 2009; 30: 1424-1432.
- 16) McCart AE, Vickaryous NK, Silver A: Apc mice: models, modifiers and mutants. Pathol Res Pract. 2008; 204, 479-490.
- 17) Abnet CC, Freedman ND, Hu N, Wang Z, Yu K, Shu XO, Yuan JM, Zheng W, Dawsey SM, Dong LM, Lee MP, Ding T, Qiao YL, Gao YT, Koh WP, Xiang YB, Tang ZZ, Fan JH, Wang C, Wheeler W, Gail MH, Yeager M, Yuenger J, Hutchinson A, Jacobs KB, Giffen CA, Burdett L, Fraumeni JF Jr, Tucker MA, Chow WH, Goldstein AM, Chanock SJ, Taylor PR: A shared susceptibility locus in *PLCE1* at 10q23 for gastric adenocarcinoma and esophageal squamous cell carcinoma. Nat Genet. 2010; 42: 764-767.
- 18) Wang LD, Zhou FY, Li XM, Sun LD, Song X, Jin Y, Li JM, Kong GQ, Qi H, Cui J, Zhang LQ, Yang JZ, Li JL, Li XC, Ren JL, Liu ZC, Gao WJ, Yuan L, Wei W, Zhang YR, Wang WP, Sheyhidin I, Li F, Chen BP, Ren SW, Liu B, Li D, Ku JW, Fan ZM, Zhou SL, Guo ZG, Zhao XK, Liu N, Ai YH, Shen FF, Cui WY, Song S, Guo T, Huang J, Yuan C, Huang J, Wu Y, Yue WB, Feng CW, Li HL, Wang Y, Tian JY, Lu Y, Yuan Y, Zhu WL, Liu M, Fu WJ, Yang X, Wang HJ, Han SL, Chen J, Han M, Wang HY, Zhang P, Li XM, Dong JC, Xing GL, Wang R, Guo M, Chang ZW, Liu HL, Guo L, Yuan ZQ, Liu H, Lu Q, Yang LQ, Zhu FG, Yang XF, Feng XS, Wang Z, Li Y, Gao SG, Qige Q, Bai LT, Yang WJ, Lei GY, Shen ZY, Chen LQ, Li EM, Xu LY, Wu ZY, Cao WK, Wang JP, Bao ZQ, Chen JL, Ding GC, Zhuang X, Zhou YF, Zheng HF, Zhang Z, Zuo XB, Dong ZM, Fan DM, He X, Wang J, Zhou Q, Zhang QX, Jiao XY, Lian SY, Ji AF, Lu XM, Wang JS, Chang FB, Lu CD, Chen ZG, Miao JJ, Fan ZL, Lin RB, Liu TJ, Wei JC, Kong QP, Lan Y, Fan YJ, Gao FS, Wang TY, Xie D, Chen SQ, Yang WC, Hong JY, Wang L, Qiu SL, Cai ZM, Zhang XJ: Genome-wide association study of esophageal squamous cell carcinoma in Chinese subjects identifies susceptibility loci at PLCE1 and C20orf54. Nat Genet. 2010; 42: 759-763.